

Zakład Fizjologii Roślin. Instytut Biologii. Uniwersytet Marii Curie-Skłodowskiej
Kierownik: prof. dr hab. Tadeusz Baszyński

Adam PASZEWSKI, Jan JAROSZ

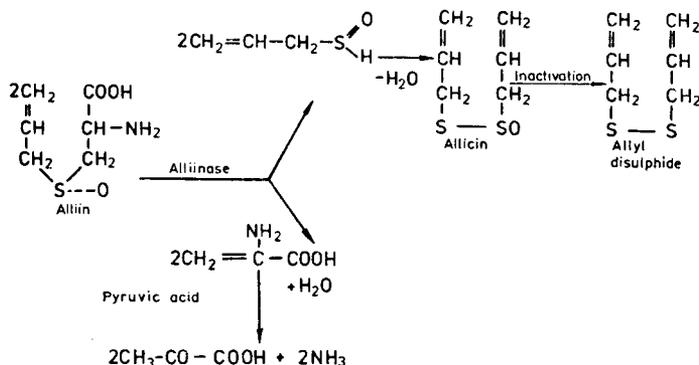
Antimicrobial Action of Garlic (*Allium sativum* L.) and Garlic Preparations Produced in Poland

Przeciwdrobnoustrojowe działanie czosnku (*Allium sativum* L.) i preparatów czosnkowych produkowanych w Polsce

Противмикробное действие чеснока (*Allium sativum* L.) и чесночных препаратов вырабатываемых в Польше

The ethereal substances with antimicrobial action produced by higher plants were named phytoncides by Tokin (14). Many of them reveal a broad spectrum of antibiotic activity: they are fungicidal and bactericidal against Gram-positive as well as Gram-negative bacteria and also protozoocidal. At low concentrations phytoncides inhibit the growth of various pathogenic yeasts and moulds. They play a significant role in the antibiotic defence mechanism of higher plants.

The curative action of garlic (*Allium sativum* L.) has been known for a long time in folk medicine, but it was not until 1945 that Cavallito and Bailey (1) described the active substance which they isolated from garlic bulbs and named



Schema 1. Enzymatic conversion of alliin into allicin (4) and its inactivation in the form of diallyldisulphide

it allicin. Allicin proved to be an allyl ester of thiosulphinic acid (2, 9) and it is formed from a biologically inactive precursor of S-allylcysteine sulphoxide named alliin under the influence of alliinase, the enzyme found in garlic tissues (1, 3, 11, 12). This antibiotic is an unstable compound and easily turns into a biologically inactive component, diallyldisulphide (4, 6, 11, 12). The scheme of enzymatic conversion alliin into allicin and the inactivation of the antibiotic in the form of diallyldisulphide is shown in schema 1.

Several authors (13, 17) stated that allicin is an inhibitor of -SH group of respiratory enzymes. Among them xantine oxidase, succinic dehydrogenase and triose phosphate dehydrogenase are most strongly inhibited. The -SO-S-grouping is essential for bactericidal action of allicin (9), and this grouping is also essential for the inhibition of -SH-enzymes, while -S-S-, and -SO-groupings are not effective (17).

It has been reported by several investigators (5, 6, 10, 15, 16) that garlic juice is a very potent antimicrobial agent both against bacteria and *Candida albicans*, and against other pathogenic yeasts. As stated by Tynecka and Skwarek (16), garlic juice did not exhibit any toxic effect against the tissue cultures of chick embryo fibroblasts or monkey kidney cells. At the same concentration it inhibited the growth of the infecting yeasts of *C. albicans*.

Considering the strong antimicrobial action and the broad spectrum of antibiotic activity of allicin, several attempts were made in Poland and abroad to make a medicine out of garlic. At present in Poland, Poznańskie Zakłady Zielarskie Herbapol produce two preparations from this raw material: Allio stabil as an ethanolic extract from fresh garlic bulbs containing about 0.05% of ethereal sulphur compounds, and Allio phil-lyophilizate of garlic prepared in pills.

The authors of this paper estimate critically the biological action of these preparations as compared with the antimicrobial action of fresh garlic bulbs.

MATERIALS AND METHODS

1. Garlic bulbs (*Allium sativum* L.) from 1976 crops.
2. Allio phil (Contents: *Allium sativum lyoph.* 0.20, *Massa tab. et drag.* ad 0.6 g), series 1711175, expiry date 15.II.1978.
3. Allio stabil (ethanolic garlic juice), series 031275, without expiry date.
4. Federative Republic of Germany preparation "Sanhelios 333" containing extracts from *Allium sativum*, *Viscum album*, *Crataegus* sp.

Preparation of pulp and active garlic juice. After removing the outer parts, the garlic bulbs were frozen in solid CO₂, then crushed thoroughly and homogenized at 3000 rpm for 5 min. The garlic juice accurately pressed through a sterile gauze was sterilized through a bacteriological funnel Schot G-4, and then the filtrate was preincubated at 37°C for 30 minutes in order to convert alliin into allicin, the antibiologically active substance. Several microbiological methods were used in order to compare the antimicrobial action of garlic, Allio phil and Allio satbil. Allio phil (*A. sativum lyoph.*) pills without coats were ground into powder and extracted with 96% ethanol or water (100 mg of the preparation per 1 ml) for 2 hrs.

Paper disc agar diffusion method. In order to determine the antimicrobial spectrum of juice and pulp from garlic and preparations, the paper disc agar plate assay was used. Each Petri plate contained 10 ml of optimal agar

medium and 0.025 ml of microbial culture from the exponential growth phase. On each disc (13 mm diameter) 100 mg of pulp was put or 0.05 ml (one drop) of garlic juice, which diffused into agar medium and produced a growth inhibition zone. Approximately 0.05 ml of Allio-stabil or ethanolic extract of Allio-phil were poured on separate paper discs.

Two-fold agar (and broth) dilution methods. The garlic juice or solutions of garlic preparations were diluted in 30% ethanol or water and poured in amounts 1 ml into 9 ml optimal agar or liquid medium. In this way a number of plates were prepared with a decreasing concentration of the antibiotic in the assay medium. The plates were inoculated with 0.5×10^6 CFU of the test organism per 1 ml of the medium and incubated at 35°C. The drop-count test and cup-plate method were also used for these comparisons.

Maximal Inhibitory Dilution (MID) was defined as the amount of antibiotic contained in the greatest dilution, which completely inhibited the growth of the test organism at 35°C for 48 hrs incubation. Maximal Biocidal Dilution (MBD) was defined as a maximal dilution of the antibiotic which completely inhibited the growth of the test organisms for 15 days incubation. After this time the viable cell units of the test organisms usually were not ascertained, as demonstrated by plate count test.

RESULTS AND DISCUSSION

The antimicrobial action of garlic determined by the paper disc agar plate assay was strong and included a wide range of the tested micro-organisms, Gram-positive and Gram-negative bacteria, acid-fast bacteria as well as yeasts and moulds. Under the same conditions, the action of Allio-stabil was light and became visible only in the case of organisms especially sensitive to allicin. The action of Allio-phil was similar and in the trace amounts it inhibited *Corynebacterium equi*, *Schizosaccharomyces octosporus* and *Aspergillus niger*, as shown in Table 1.

The MID values of garlic juice and garlic preparations assayed by the twofold agar dilution method for 4 test organisms (*Bacillus subtilis* ATCC 6633, *Sarcina lutea* R-262, *Proteus vulgaris* OX₁₉ and *Candida arborea*) are shown in Table 2. In comparison with Allio-phil and Allio-stabil also in this method garlic juice had a stronger effect against all four organisms tested. From the above data, it is obvious that Allio-phil action was two times or even weaker than the Allio-stabil action, and MID values determined for garlic juice were in some cases more than eight times higher than those of the Allio-stabil action. We have not included in this comparison the antimicrobial action of pulp which contains (as Table 1 indicates) a considerable amount of allicin activity.

The garlic juice had a strong antibiotic action against *S. lutea*, *Pseudomonas aeruginosa* and *Escherichia coli* by the drop-count test in the semi-liquid (0.4%) agar medium and by a cup-plate agar diffusion assay while under the same conditions no trace of the preparations' action was seen.

Table 1. Comparison of antimicrobial action of garlic juice and pulp, and garlic preparations by paper disc agar plate assay procedure

Microorganisms	Medium	Garlic juice	Garlic pulp	Alliophil	Alliostabil	
		Diameter inhibition zone in mm				
<i>Bacillus subtilis</i> ATCC 6633		43	66	—	Trace	
<i>Bacillus cereus</i> ATCC 8145		47	69	—	17	
<i>Bacillus lentus</i> IP 5286		40	63	—	Trace	
<i>Bacillus brevis</i> IP 5275		42	54	—	Trace	
<i>Escherichia coli</i> ROW	Nutrient agar	32	45	—	—	
<i>Proteus vulgaris</i> OX ₁₉		30	39	—	—	
<i>Pseudomonas</i> <i>aeruginosa</i> H ₃		20	26	—	—	
<i>Salmonella pullorum</i>		32	43	—	—	
<i>Sarcina lutea</i>		36	51	—	—	
<i>Micrococcus</i> <i>lysodeikticus</i>		36	52	—	—	
<i>Staphylococcus</i> <i>aureus</i> 209-P		43	61	—	—	
<i>Staphylococcus albus</i>		48	67	—	—	
<i>Corynebacterium xerosis</i>		Loeffler agar	60	80	—	—
<i>Corynebacterium equi</i>			52	74	20	20
<i>Mycobacterium</i> sp. ATCC 607	Sauton agar	46	73	—	19	
<i>Mycobacterium</i> <i>smegmatis</i>		42	69	—	Trace	
<i>Mycobacterium</i> <i>butyricum</i>		40	68	—	Trace	
<i>Mycobacterium</i> sp. ATCC 279		40	63	—	17	
<i>Saccharomyces</i> <i>cerevisiae</i>	Malt- -agar	41	56	—	—	
<i>Saccharomyces</i> <i>carlsbergensis</i>		39	48	—	—	
<i>Candida arborea</i>		51	82	—	Trace	
<i>Rhodotorula rubra</i>		38	57	—	19	
<i>Schizosaccharomyces</i> <i>octosporus</i>		53	100	18	20	
<i>Aspergillus niger</i>		59	100	20	25	

Table 2. Comparative MID's of garlic juice, Alliophil (ethanolic extract) and Alliostabil assayed by the twofold agar dilution method

Organism tested	MID, per cent basic solution		
	Garlic juice	Alliophil	Alliostabil
<i>Bacillus subtilis</i> ATCC 6633	0.306	5.0	2.5
<i>Sarcina lutea</i> R-262	0.306—0.612	7.5	2.5
<i>Proteus vulgaris</i> OX ₁₉	0.612	7.5	3.75
<i>Candida arborea</i>	0.153	1.25—2.5	1.25

Inoculum size; approx. 0.5×10^6 CFU/ml. The MID values for *C. arborea* determined in malt-agar (7° Blg), other organisms tested in nutrient agar 1.4% at 35° for 24 hours incubation.

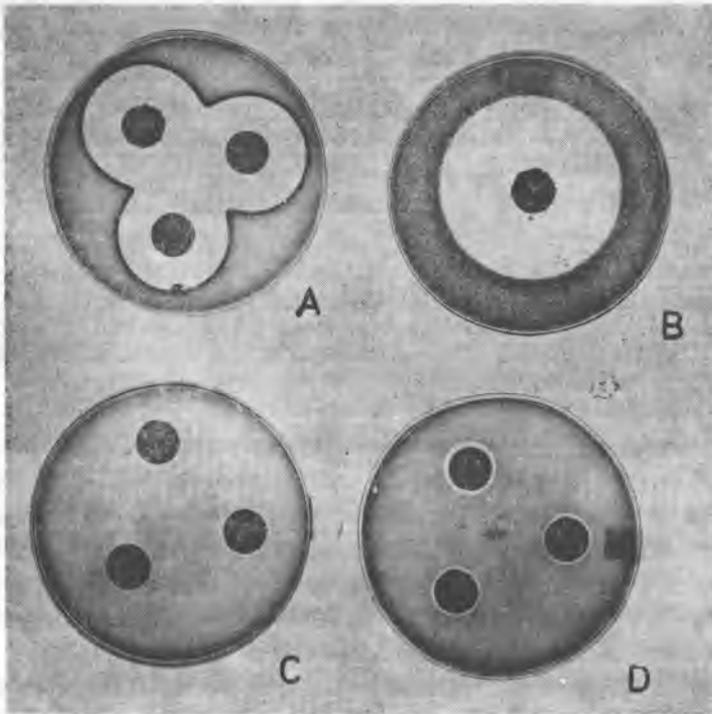


Fig. 1. *Rhodotorula rubra* growth inhibition zones by the paper disc agar diffusion assay procedure; A — garlic juice, B — garlic pulp, C — Alliophil, D — Alliostabil

The results of the assays of the antimicrobial action of garlic juice and garlic preparation using the paper disc agar plate method and the twofold agar dilution method suggested that the diffusive methods are not entirely useful for measuring the antibiotic action of alliin because it seems that this antibiotic after isolation from garlic tissues diffuses weakly in agar media. In the case of the paper disc assay only microorganisms very susceptible to the alliin action were inhibited, mainly yeasts and moulds for which the values of MIC are the lowest, and the amount of antibiotic which diffused from the paper disc to agar medium could reach this level (Fig. 1).

Due to that, the activity of garlic and garlic preparations was assayed by the twofold broth dilution technique against *B. subtilis* ATCC 6633 as a test organism used for the quantitative estimation of alliinase (8), the enzyme responsible for the conversion of the parent form into alliin, a form biologically active. The action of preparations estimated by this method was much higher than those estimated by the diffusive methods and close to action of garlic (strong pulp action was not compared), but transient and bacteriostatic, bactericidal only in higher concentrations, as

Table 3. Biostatic and biocidal action of garlic juice and garlic preparations determined by the twofold broth dilution assay procedure for *Bacillus subtilis* ATCC 6633 as a test organism

Preparation	Incubation period (days)	2	10	15	Ratio of MBD/MID
		MID, per cent basic solution			
Garlic juice		0.153	0.306	0.306	2
Alliostabil		0.306	0.612	1.25	4
Alliophil					
ethanolic solution		1.25	3.75	7.50	6
aqueous solution	10	10	10	>10	—
Sanhelios 333	10	10	>10	>10	—

Assay medium and pH value; nutrient broth, pH 6.5; — not determined. The MBO values (maximal biocidal dilution); lowest allicin concentration in the garlic solution resulting in no visible growth after 15 days incubation.

shown in Table 3. For the garlic juice the ratio of MBD/MID was 2, and for Alliostabil and Alliophil were 4 and 6, respectively. Generally, the antimicrobial action of garlic by the twofold broth dilution method was two times as strong as that of Alliostabil, and eight times as strong as that of ethanolic solutions of Alliophil. However, the antibiotic activity of Sanhelios 333 was very weak, only at 10% concentration the growth of *B. subtilis* was completely inhibited for 3 days.

Allicin is an oil with a characteristic garlic scent, soluble in benzene, ethanol and ether; its solubility in water at 10° comes to 2.5%. In aqueous solutions or in dry base allicin becomes inactive at room temperature after two days, forming a mucilaginous liquid (7). From the above findings and comparisons, it is reasonable to conclude that the comparatively weak action of Alliophil and Alliostabil was connected with the insignificant diffusion of this antibiotic to solid agar media (when they bioactivity were assayed by diffusive methods) as with the inactivation of allicin after isolating this antibiotic from garlic tissues. Similarly, K a b e l i k (6) has been reported that aqueous, ethanolic and acidic extracts from garlic bulbs possessed a weak antimicrobial activity, however, lyophilizate of garlic contains a considerable amount of allicin activity (5). In fresh garlic juice there is both a precursor of this antibiotic and an active enzyme alliinase capable of transforming alliin into allicin and this explains the strong bactericidal action of garlic bulbs.

The authors of this communication do not deny the antimicrobial action of Alliophil and Alliostabil, but they call in question the use of these as remedy. They think garlic bulbs are more powerful. All our experiments were performed *in vitro*.

REFERENCES

1. Cavallito C. J., Bailey J. H.: J. Am. Chem. Soc. **66**, 1950—1952, 1944.
2. Cavallito C. J., Buck J. S., Suter C. M.: J. Am. Chem. Soc. **66**, 1952—1954, 1944.
3. Cavallito C. J., Bailey J. H., Buck J. S.: J. Am. Chem. Soc. **67**, 1032—1033, 1945.
4. Hörheimer L., Wagner H., Seitz M., Vejdeck Z.: Pharmazie **23**, 462—467, 1968.
5. Jeżowa L., Rafiński T., Wrociński T.: Herba Polon. **1**, 1—13, 1966.
6. Kabelik J.: Pharmazie, **25**, 266—270, 1970.
7. Korzybski T., Kowszyk - Gindifer Z., Kuryłowicz W.: Antibiotics. Pergamon Press, Oxford 1967.
8. Rao R. R., Krishna M. C., cited after Korzybski et al. (7).
9. Small L. D., Bailey J. H., Cavallito C. J.: J. Am. Chem. Soc. **69**, 1710—1713, 1947.
10. Spivak M. Y., Argubayeva H. A., Konoshenko M. F.: Antibiotiki **8**, 832—833, 1963.
11. Stoll A., Seebeck E.: Helv. Chim. Acta **31**, 189—210, 1948.
12. Stoll A., Seebeck E.: Helv. Chim. Acta **32**, 197—205, 1949.
13. Szymona M.: Acta Microbiol. Polon. **1**, 5—23, 1952.
14. Tokin B. P.: Fitocydy, Moskwa 1951.
15. Tynecka Z., Goś Z.: Acta Microbiol. Polon. **5**, 51—62, 1973.
16. Tynecka Z., Skwarek T.: Farmacja Polska **30**, 531—538, 1974.
17. Willis E. D.: Biochem. J. **63**, 514—520, 1956.

Otrzymano 8 II 1978.

STRESZCZENIE

Przy użyciu szeregu metod oznaczania aktywności antybiotyków porównano przeciwdrobnoustrojowe działanie czosnku (soku i miazgi) z aktywnością preparatów czosnkowych produkowanych w Polsce. Działanie Alliofilu (przygotowanego wg podanej w Metodach procedury) i Alliofilu było przejściowe, biostatyczne, a przede wszystkim słabsze od działania soku czosnkowego, szczególnie przy zastosowaniu dyfuzyjnych metod testowania. Stosunek dawki bójczej do biostatycznej MBC/MIC wyznaczony dla soku z czosnku, Alliofilu i Alliofilu, wynoszący odpowiednio 2, 4 i 6 wskazuje, że allicyna po wyizolowaniu z tkanek czosnku i odwodnieniu ulega w dużym stopniu inaktywacji oraz słabo dyfunduje do stałych podłoży agarowych. Natomiast sok czosnkowy zawierający allinę — prekursor allicyny oraz allinazę, enzym odpowiedzialny za enzymatyczną konwersję prekursora do formy aktywnej biologicznie, okazał się silnym, o szerokim spektrum działania, środkiem przeciwdrobnoustrojowym.

РЕЗЮМЕ

При использовании (употреблении) ряда методов определения активности антибиотиков сравнено противомикроорганизмовое действие чеснока (сока и мезги) с активностью чесночных препаратов, вырабатываемых в Польше. Действие

Аллиофила и Аллиостабила было кратковременное, биостатичное, а прежде всего значительно слабее, чем действие чесночного сока, особенно при применении диффузионных методов определения. Отношение убийственной дозы к биостатичной МВС/МИС, назначенное для чесночного сока, Аллиостабила и Аллиофила, составляющее (равняющееся) соответственно 2, 4 и 6 указывает, что аллицина после изолирования из ткани чеснока и водоотлива подвергается в большой степени инаktivации, как также слабо диффундует к постоянным агаровым материнским породам. Чесночный сок же, содержащий аллину — прекурсор аллицины, а также аллиназу, фермент ответственный за энзиматичную конверсию прекурсора до биологически активной формы, оказался сильным, в широком спектре действия противомикроорганизмовым средством.