

Faculty of Chemistry
Department of Organic Chemistry and Technology

Stanisław DACKA, Zofia ZIMIŃSKA*

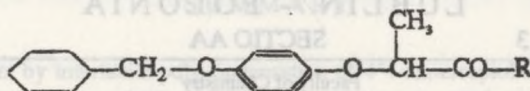
Synthesis and Biological Properties of Derivatives of α -(4-Cyclohexylmethoxyphenoxy) Propionic Acid

Synteza oraz właściwości biologiczne pochodnych
kwasu α -(4-cykloheksylometoksyfenoksy)-propionowego

Lately some reports concerning the synthesis as well as biological activity of derivatives of propionic acid have appeared. These compounds show bactericidal, fungicidal and herbicidal activities. Of these compounds derivatives of aryloxyphenoxypropionic acid reveal good phytocidal activity on monocotyledonous plants [1–5]. For this reason it seems interesting for us to find a method for synthesis of α -(4-cyclohexylmethoxyphenoxy) propionic acid unknown so far and some of its derivatives as well as to determine their biological activity.

The starting compound was α -(4-hydroxyphenoxy) propionic acid [6] obtained in the reaction of methyl α -bromopropionate and 4-methoxyphenol. This compound with cyclohexylmethyl bromide in the presence of sodium ethylate in the ethanol solution results in α -(4-cyclohexylmethoxyphenoxy) propionic acid (1). Phenylphenacyl ester was prepared by heating the compound (1) with 4-phenylphenacyl bromide in a weakly alkaline medium. Acid (1) was transformed next into acid amides in the reaction of acid chloride with the corresponding amines.

* Instytut Przemysłu Organicznego (Warszawa) – Institute of Organic Industry (Warsaw).



- | | |
|---|---|
| 1. R=OH | 9. R=NHCH ₂ CH(OH)CH ₃ |
| 2. R=OCH ₂ COC ₆ H ₄ C ₆ H ₅ (4) | 10. R=NHCH ₂ C ₆ H ₅ |
| 3. R=NH ₂ | 11. R=NHC ₆ H ₅ |
| 4. R=NHCH ₃ | 12. R=NHC ₆ H ₄ OCH ₃ (3) |
| 5. R=NHC ₂ H ₅ | 13. R=NHC ₆ H ₄ Br (4) |
| 6. R=NH(CH ₂) ₃ CH ₃ | 14. R=NHC ₆ H ₄ CH ₃ (4) |
| 7. R=NH(CH ₂) ₄ CH ₃ | 15. R=NHCH ₂ C ₄ H ₉ O (2) |
| 8. R=NHCH ₂ CH ₂ CH(CH ₃) ₂ | |

Tab. 1. Biological activity of derivatives of α -(4-cyclohexylmethoxyphenoxy)propionic acid

Compound No	Dose in ppm for one person or concentration in % causing over 90% of mortality		Blocking concentration [ppm]					% paresis concentration [1000 ppm]
	Housefly	Spider mite %	<i>Alternaria tenuis</i>	<i>Botritis cinerea</i>	<i>Rhizoctonia solani</i>	<i>Fusarium culmorum</i>	<i>Phytophthora cactorum</i>	<i>Erysiphe graminis</i>
1	>25	>0.1	>100	>100	>200	>200	>200	100
2	>25	>0.1	>100	>100	>200	>200	>200	80
3	>25	>0.1	>100	>100	>200	>200	>200	100
4	>25	>0.1	>100	>100	>200	>200	>200	46
5	>0.1%	>0.1	>100	>100	>200	>200	>200	27
6	>0.1%	>0.1	>100	>100	>200	>200	>200	33
7	>0.1%	>0.1	>100	>100	>200	>200	>200	22
8	>0.1%	>0.1	>100	>100	>200	>200	>200	35
9	>0.1%	>0.1	>100	>100	200	200	200	41
10	>0.1%	>0.1	>100	>100	>200	>200	>200	17
11	>25	>0.1	>100	>100	>200	>200	>200	100
12	>0.1%	>0.1	>100	>100	>200	>200	>200	20
13	>0.1%	>0.1	>100	>100	>200	>200	>200	35
14	>0.1%	>0.1	>100	>100	>200	>200	>200	56
15	>0.1%	>0.1	>100	>100	>200	>200	>200	19

The above-mentioned compounds have been tested for insecticidal, spidericidal, fungicidal and phytocidal activities according to the methods worked out for the screening test [7]. Investigations have been carried out with the use of bioindicators: housefly (*Musca domestica*) and spider mite (*Tetranychus urticae*). In the

experiments 1% and 0.1% dose solutions of the tested compounds were used. After 48h the lethality evaluation of the tested bioindicators was performed. Fungicidal activity for the fungi: *Alternaria tenuis*, *Botrytis cinerea*, *Rhizoctonia solani*, *Fusarium culmorum* and *Phytophthora cactorum* was studied in the tests *in vitro* as well as *in vivo* on living plants coated with spores of *Erysiphe graminis*. Phytocidal activity in pre- and post-emergence applications with the use of 10 species of plants as indicators: oat, flax, buckwheat, beet, cucumber, corn, mustard, pea, bean and ryegrass was studied. Concentrations of these compounds were chosen so that the dose would equal to 5 kg/ha.

The obtained compounds do not reveal any activity either to housefly or spider mite. Fungicidal activity towards *Erysiphe graminis* was observed in most compounds, where compounds (7), (10), (12), and (15) were characterized by good activity and the remaining – except for (1), (2), (3) and (11) – mean. Compound (9) reveals the mean fungicidal activity for the pathogens: *Rhizoctonia solani*, *Fusarium culmorum* and *Phytophthora cactorum*. The investigated compounds do not reveal phytocidal activity except α -(4-cyclohexylmethoxyphenoxy) propionic acid, the activity of which is mean for the tested plants.

EXPERIMENTAL

IR spectra were recorded in KBr discs with a FT 1725X Perkin Elmer spectrophotometer. ^1H NMR spectra were determined using BF 567A Tesla 100 spectrometer with TMS as an internal standard.

1. α -(4-Cyclohexylmethoxyphenoxy) propionic acid

A sample of 56 g (0.3 mole) of α -(4-hydroxyphenoxy) propionic acid and 55 g (0.31 mole) of cyclohexylmethyl bromide was added to the solution of sodium ethylate (prepared from 14.2 g (0.62 mole) of sodium and 250 cm³ of absolute alcohol) and was refluxed for 5 hours. The precipitate of salt was filtered and crystallized from water (100 cm³). The salt was dissolved in water and acidified with HCl. The separated acid was filtered and crystallized from benzene-cyclohexane solution (1:2) (300 cm³). Plates m.p. 129–130°C. Yield 60 g (72%).

Analysis:

For $\text{C}_{16}\text{H}_{22}\text{O}_4$ (278.36) calcd : 69.04% C; 7.94% H;
found : 69.38% C; 8.27% H.

IR [cm⁻¹] 1370, 1450, 1470 δCH_3 , CH_2 ; 2850 $\nu_{\text{as}}\text{CH}_2$, CH_3 ; 2920, 2937 $\nu_{\text{as}}\text{CH}_2$; 2980 $\nu_{\text{as}}\text{CH}_3$; 818, 840, 850, 1025, 1035, 1050, 1092, 1140, 1215, 1225 $\delta\text{C}_{\text{Ar}}$ —H

(subst. 1.4); 1510, 1627 $\nu_{C_{Ar}=C_{Ar}}$; 943 $\nu_{OH(COOH)}$; 3430 $\nu_{OH(COOH)}$; 1290 $\nu_{C-O(COOH)}$; 1735 $\nu_{C=O(COOH)}$.

1H NMR [δ ppm] ($CDCl_3$); 0.75–2 m 11H(C_6H_{11}); 1.52 d, $J=6.6$ Hz, 3H(CH_3); 3.70 d $J=5.6$ Hz, 2H(CH_2O); 4.70 q $J=6.6$ Hz, 1H(CHO); 6.71 s 4H(OC_6H_4O); 10.75 s 1H(COOH).

2. 4-Phenylphenacyl ester of α -(4-cyclohexylmethoxyphenoxy) propionic acid

4 g (0.014 mole) of acid (1) were neutralized with 3% NaOH to pH 7.2. To this solution 3.5 g (0.014 mole) of 4-phenylphenacyl bromide in 60 cm³ of ethanol were added and the mixture was refluxed for 1 h. After cooling the precipitate was filtered and crystallized from methanol. Plates m.p. 106–107°C. Yield 5.5 g (83%).

Analysis:

For $C_{30}H_{32}O_5$ (472.58) calcd: 76.25% C; 6.83% H;

found: 76.52% C; 6.62% H.

IR [cm^{-1}] 1762 $\nu_{C=O}$; 1191 ν_{C-O-C} .

1H NMR [δ ppm] ($CDCl_3$) 0.94–1.9 m 11H(C_6H_{11}); 1.74 d $J=7$ Hz 3H(CH_3-CH); 3.68 d $J=6.1$ Hz 2H(CH_2O); 4.85 q $J=7$ Hz 1H(CHO); 5.42 s 2H(CH_2-CO); 6.76–7.99 m 13H (aromatic).

Preparation of *N*-substituted amides

Method A

6 g (0.021 mole) of acid (1) and 25 cm³ of thionyl chloride were refluxed until the precipitate was almost completely dissolved (1 h). Thionyl chloride was removed under the reduced pressure. 50 cm³ of 20% aqueous solution of amine were added to the residue. The precipitate was filtered and washed with water.

3. α -(4-Cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 154–156°C (benzene-cyclohexane (1:1)). Yield 5.5 g (92%)

Analysis:

For $C_{16}H_{23}NO_3$ (277.36) calcd : 5.05% N;

found: 5.05% N.

IR [cm^{-1}] 1600 $\delta N-H$; 3185 $\nu_s N-H$; 3390 $\nu_{as} N-H$; 1670 $\nu_{C=O}$.

4. *N*-Methyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 94.5–95.5°C (cyclohexane). Yield 5.3 g (85%).

Analysis:

For $C_{17}H_{25}NO_3$ (2291.39) calcd: 4.81% N;

found: 5.17% N.

IR [cm^{-1}] 1544 $\delta\text{N—H}$; 3328 $\nu\text{N—H}$; 1661 $\nu\text{C=O}$

$^1\text{H NMR}$ [δppm] (CDCl_3) 0.96–1.9 m 11H(C_6H_{11}); 1.52 d $J=6.6$ Hz 3H($\text{CH}_3\text{—CH}$); 3.4 d $J=7$ Hz 3H($\text{CH}_3\text{—NH}$); 3.7 d $J=5.6$ Hz 2H(CH_2O); 4.54 q $J=6.6$ Hz 1H(CHO); 6.57 1H(NH); 6.81 s 4H($\text{OC}_6\text{H}_4\text{O}$).

5. *N*-Ethyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 85.5–86°C (benzene-hexane (1:5)). Yield 5.6 g (85%).

Analysis:

For $\text{C}_{18}\text{H}_{27}\text{NO}_3$ (305.42) calcd : 4.69% N;
found: 4.55% N.

IR [cm^{-1}] 1539 $\delta\text{N—H}$; 3325 $\nu\text{N—H}$; 1654 $\nu\text{C=O}$.

$^1\text{H NMR}$ [δppm] (CDCl_3) 0.95–1.9 m 11H(C_6H_{11}); 1.12 t $J=7$ Hz 3H(CH_3CH_2); 1.52 d $J=6.6$ Hz 3H(CH_3CH); 3.3 q $J=7$ Hz 2H(CH_2N); 3.7 d $J=6.1$ Hz 2H(CH_2O); 4.54 q $J=6.6$ Hz 1H(CHO); 6.53 1H(NH); 6.81 s 4H($\text{OC}_6\text{H}_4\text{O}$).

Method B

6 g (0.021 mole) of acid (1) were converted into acid chloride as in method A. The reaction product was heated (1 h) with 0.06 mole of amine in 120 cm^3 of dry benzene. The solvent was removed under the reduced pressure. The precipitate washed with 5% HCl and water.

6. *N*-Butyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Lumps m.p. 73–73.5°C (hexane). Yield 5.7 g (81%).

Analysis:

For $\text{C}_{20}\text{H}_{31}\text{NO}_3$ (333.47) calcd: 4.20% N;
found: 4.06% N.

IR [cm^{-1}] 1542 $\delta\text{N—H}$; 3318 $\nu\text{N—H}$; 1657 $\nu\text{C=O}$.

$^1\text{H NMR}$ [δppm] (CDCl_3) 0.88–1.9 m 18H(C_6H_{11} , $\text{CH}_2\text{CH}_2\text{CH}_3$); 1.52 d $J=6.6$ Hz 3H(CH_3CH); 3.27 q $J=6.6$ Hz 2H($\text{CH}_2\text{—N}$); 3.69 d $J=5.6$ Hz 2H(CH_2O); 4.54 q $J=6.6$ Hz 1H(CHO); 6.53 1H(NH); 6.81 s 4H($\text{OC}_6\text{H}_4\text{O}$).

7. *N*-Pentyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Lumps m.p. 65.5–66.5°C (hexane). Yield 6.3 g (84%).

Analysis:

For $\text{C}_{21}\text{H}_{33}\text{NO}_3$ (347.5) calcd : 4.03% N;
found: 4.07% N.

IR [cm^{-1}] 1542 $\delta\text{N—H}$; 3321 $\nu\text{N—H}$; 1657 $\nu\text{C=O}$.

^1H NMR [δppm](CDCl_3) 0.79–1.86 m 20H(C_6H_{11} , $\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_3$); 1.53 d $J=7$ Hz 3H($\text{CH}_3\text{—CH}$); 3.25 q $J=6.6$ Hz 2H($\text{CH}_2\text{—N}$); 3.68 d $J=5.6$ Hz 2H(CH_2O); 4.54 q $J=6.6$ Hz 1H(CHO); 6.51 1H(NH); 6.81 s 4H($\text{OC}_6\text{H}_4\text{O}$).

8. *N*-3-Methylbutyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 75–76°C (hexane). Yield 5.9 g (79%).

Analysis:

For $\text{C}_{21}\text{H}_{33}\text{NO}_3$ (347.5) calcd: 4.03% N;
found: 3.75% N.

IR [cm^{-1}] 1540 $\delta\text{N—H}$; 3335 $\nu\text{N—H}$; 1660 $\nu\text{C=O}$.

^1H NMR [δppm](CDCl_3) 0.88 d $J=6.1$ Hz 6H($(\text{CH}_3)_2\text{CH}$); 0.91–1.9 m 11H(C_6H_{11}); 1.52 d $J=6.6$ Hz 3H(CH_3CH); 3.28 q $J=7$ Hz 2H(CH_2NH); 3.69 d $J=6.1$ Hz 2H(CH_2O); 4.54 q $J=6.1$ Hz 1H(CHO); 6.49 1H(NH); 6.81 s 4H($\text{OC}_6\text{H}_4\text{O}$).

9. *N*-2-Propanolo α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 85–86°C (hexane). Yield 6.2 g (86%).

Analysis:

For $\text{C}_{19}\text{H}_{29}\text{NO}_4$ (335.44) calcd: 4.17% N;
found: 4.03% N.

IR [cm^{-1}] 1547 $\delta\text{N—H}$; 3334 $\nu\text{N—H}$ and OH; 1660 $\nu\text{C=O}$.

^1H NMR [δppm](CDCl_3) 0.94–2.15 m 14H(C_6H_{11} , CH_3CHOH); 1.53 d $J=6.6$ Hz 3H(CH_3CHO); 2.98–3.2 m 2H(CH_2NH); 3.69 d $J=5.6$ Hz 2H(CH_2O); 3.86 1H(OH); 4.56 q $J=7$ Hz 1H(CHO); 6.91 s 4H($\text{OC}_6\text{H}_4\text{O}$); 7.00 1H(NH).

10. *N*-Benzyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 91–91.5°C (hexane). Yield 6.5 g (82%).

Analysis:

For $\text{C}_{23}\text{H}_{29}\text{NO}_3$ (367.5) calcd: 3.91% N;
found: 3.47% N.

IR [cm^{-1}] 1535 $\delta\text{N—H}$; 3340 $\nu\text{N—H}$; 1657 $\nu\text{C=O}$.

^1H NMR [δppm](CDCl_3) 0.95–1.9 m 11H(C_6H_{11}); 1.56 d $J=6.6$ Hz 3H(CH_3CH); 3.69 d $J=6.1$ Hz 2H(CH_2O); 4.46 d $J=5.6$ Hz 2H(CH_2N); 4.62 q $J=6.6$ Hz 1H(CHO); 6.8 s 4H($\text{OC}_6\text{H}_4\text{O}$); 6.88–7.32 m 6H(C_6H_5 , NH).

11. *N*-Phenyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 109.5–110°C (ethanol). Yield 6.7 g (88%).

Analysis:

For $\text{C}_{22}\text{H}_{27}\text{NO}_3$ (353.47) calcd: 3.96% N;
found: 4.20% N.

IR [cm^{-1}] 1530 $\delta\text{N—H}$; 3308 $\nu\text{N—H}$; 1663 $\nu\text{C=O}$.

12. *N*-3-Methoxyphenyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Plates m.p. 87.5–88.5°C (hexane). Yield 7.2 g (87%).

Analysis:

For $\text{C}_{23}\text{H}_{29}\text{NO}_4$ (383.49) calcd: 3.65% N;
found: 3.52% N.

IR [cm^{-1}] 1527 $\delta\text{N—H}$; 3295 $\nu\text{N—H}$; 1661 $\nu\text{C=O}$.

^1H NMR δppm (CDCl_3) 0.93–1.87 m 11H(C_6H_{11}); 1.59 d $J=6.6$ Hz 3H(CH_3CH); 3.67 d $J=6.1$ Hz 2H(CH_2O); 3.76 s 3H(CH_3O); 4.63 q $J=6.6$ Hz 1H(CHO); 6.6–7.35 m 8H(7H aromatic, NH); 8.31 s 1H(aromatic).

13. *N*-4-Bromophenyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 113–114°C (hexene). Yield 7.8 g (84%).

Analysis:

For $\text{C}_{22}\text{H}_{26}\text{BrNO}_3$ (432.36) calcd: 3.24% N;
found: 3.40% N.

IR [cm^{-1}] 1524 $\delta\text{N—H}$; 3304 $\nu\text{N—H}$; 1672 $\nu\text{C=O}$

^1H NMR [δppm] (CDCl_3) 1.05–1.88 m 11H(C_6H_{11}); 1.6 d $J=7$ Hz 3H(CH_3CH); 3.69 d $J=5.6$ Hz 2H(CH_2O); 4.51 q $J=6.6$ Hz 1H(CHO); 6.29 1H(NH); 6.86 s 4H($\text{OC}_6\text{H}_4\text{O}$); 7.44 s 4H($\text{C}_6\text{H}_4\text{Br}$)

14. *N*-4-Methylphenyl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles m.p. 90–91°C (heptane). Yield 6.8 g (85%).

Analysis:

For $\text{C}_{23}\text{H}_{29}\text{NO}_3$ (367.49) calcd: 3.81% N;
found: 3.70% N.

IR [cm^{-1}] 1526 $\delta\text{N—H}$; 3305 $\nu\text{N—H}$; 1660 $\nu\text{C=O}$.

15. *N*-2-Furfuryl α -(4-cyclohexylmethoxyphenoxy) propionamide

Needles 83.5–84°C (hexane). Yield 6.5 g (84%).

Analysis:

For $\text{C}_{21}\text{H}_{27}\text{NO}_4$ (357.45) calcd: 3.77% N;
found: 3.91% N.

IR [cm^{-1}] 1546 $\delta\text{N—H}$; 3295 $\nu\text{N—H}$; 1665 $\nu\text{C=O}$.

^1H NMR [δppm] (CDCl_3) 0.94–1.9 m 11H(C_6H_{11}); 1.53 d $J=7$ Hz 3H(CH_3CH); 3.68 d $J=5.6$ Hz 2H(CH_2O); 4.45 d $J=5.6$ Hz 2H(CH_2N); 4.59 q $J=6.6$ Hz 1H(CHO); 6.1–6.3 m 3H(NH, 2H positions 3 and 4 in furane); 6.8 s 4H($\text{OC}_6\text{H}_4\text{O}$); 7.3 d 1H(position 5 in furane).

REFERENCES

- [1] White S. R.: *Pest. środ.*, **49** (1977).
- [2] Kenneth H. A.: *Chem. pesticid.*, **265** (1982).
- [3] Carter Ch. G., Lee D. L.: US Pat. No. 4613 357 (1986).
- [4] Fujinawa Shoji, Hashiba Isao, Suzuki Kenji: Jap. Pat. No. 61 158 947 (1986).
- [5] Rehn K., Nester J. H.: Ger Pat. 3240 805 (1984).
- [6] Lewis R. H., Housley R. J., Richards C. H., Nicholson S. J., Baker W. M., Adams S. St.: GB Pat. 916 242 (1959).
- [7] Bakuniak E., Bakuniak J.: *Pestycydy*, **12**, 4 (1978).

STRESZCZENIE

W ostatnich latach opublikowano wiele artykułów nt. syntezy oraz aktywności biologicznej pochodnych kwasu propionowego. Takie pochodne wykazują właściwości grzybobójcze, bakteriobójcze, jak również chwastobójcze. Na przykład pochodne kwasu aryloksyfenoksypropionowego wykazują silne działanie fitocydalne na rośliny jednoliścienne [1–5]. W związku z tym wydało się celowe opracowanie metody syntezy dotychczas nieznanego kwasu α -(4-cykloheksylometoksyfenoksy)-propionowego i niektórych jego pochodnych, a także zbadanie ich biologicznej aktywności.

Substratem w przeprowadzonych doświadczeniach był kwas α -(4-hydroksyfenoksy)-propionowy [6] otrzymany w reakcji α -bromopropionianu metylu z 4-metoksyfenolem. Kwas α -(4-hydroksyfenoksy)-propionowy tworzył z bromkiem cykloheksylometylowym w etanolu w obecności etanolanu sodowego kwas α -(4-cykloheksylometoksyfenoksy)-propionowy (1). Ester 4-fenylofenacylowy otrzymano przez ogrzewanie kwasu (1) z bromkiem 4-fenylofenacylowym w środowisku słabo alkalicznym. Kwas karboksylowy (1) przeprowadzono w pochodne amidowe przez działanie na chlorek kwasowy (otrzymany przez ogrzewanie kwasu (1) z chlorkiem tionylu) odpowiednimi aminami.

Wymienione związki zostały poddane badaniu, które miało wyjaśnić, czy wykazują aktywność owado- i przedziorkobójczą, grzybobójczą i fitocydalną (metodami opracowanymi dla *screening* testu) [7]. Badania przeprowadzono używając bioindykatorów: muchy domowej (*Musca domestica*) i przedziorka chmielowca (*Tetranychus urticae*). W doświadczeniach stosowano 1% i 0,1% dawkę roztworu badanego związku. Po upływie 48 h przeprowadzono ocenę śmiertelności badanych bioindykatorów. Aktywność grzybobójczą badano w testach *in vitro* dla grzybów *Alternaria tenuis*, *Botrytis cinerea*, *Rhizoctonia solani*, *Fusarium culmorum* i *Phytophthora cactorum* oraz *in vivo* na żywych roślinach pokrytych zarodnikami mączniaka właściwego (*Erysiphe graminis*). Działanie fitocydalne zbadano przedwschodowo i powschodowo z użyciem 10 gatunków roślin wskaźnikowych, takich jak owies, len, gryka, burak, ogórek, kukurydza, gorczyca, groch, fasola i rajgras, przy czym stężenia związków były tak dobrane, aby dawka wynosiła 5 kg/ha.

Badane związki nie były aktywne ani w stosunku do muchy domowej, ani do przedziorka. Nie wielką aktywność grzybobójczą wobec *Erysiphe graminis* wykazała większość połączeń: związki (7), (10), (12) i (15) wykazują działanie silne, a pozostałe – z wyjątkiem (1), (2), (3) i (11) – aktywność przeciętną. Związek (9) wykazuje przeciętną aktywność grzybobójczą wobec takich patogenów, jak *Rhizoctonia solani*, *Fusarium culmorum* i *Phytophthora cactorum*. Badane związki nie wykazują aktywności fitocydalnej (z wyjątkiem kwasu α -(4-cykloheksylometoksyfenoksy)-propionowego, który wykazuje przeciętną aktywność wobec roślin testowanych).